

# Robust Flux Balance Analysis of Metabolic Networks

Michael M. Zavlanos and A. Agung Julius

**Abstract**—Metabolic networks describe the set of biochemical reactions and regulatory interactions of metabolism that govern the phenotypical properties of a cell. Analysis of such networks is critical not only to promote biological knowledge, but also in drug discovery, where it can be used to identify and knockout the targeted pathways. Flux Balance Analysis (FBA) has been widely used to study metabolic networks. This powerful technique employs the reaction stoichiometries and reversibility constraints along with experimental measurements of phenotypical properties of the cell, e.g., biomass composition or ATP synthesis, to compute the fluxes of metabolites that are best manifested in the cell. Although FBA has been shown to satisfactorily capture cell behavior, its performance could be significantly improved if measurement uncertainty is introduced in the models. In this paper we propose Robust Flux Balance Analysis (RFBA) to determine optimal fluxes of metabolites for all phenotypical measurements in a given uncertainty set. We derive a least squares bi-criterion approximation of the uncertain problem and, using the S-procedure and tools from matrix analysis, we show that this is equivalent to a semidefinite program that can be solved optimally using available techniques. We illustrate our approach on synthetic metabolic networks and discuss the effect of regularization on the final solutions. Due to its convex nature, our approach can be applied to genome-scale networks.

## I. INTRODUCTION

Metabolic networks map the biochemical reactions in a living cell to the flow of various chemical substances in the cell, which are called metabolites. The metabolic network of an organism can be thought of as production lines in a large scale biochemical plant. It captures the totality of metabolic reactions in which chemical substances are consumed to produce metabolic products. Analysis of such networks is critical not only to promote biological knowledge, but also in drug discovery, where it can be used to identify and knockout the targeted pathways.

Metabolic Flux Balance Analysis (FBA) [1], [2] studies the feasible and optimal reaction fluxes through the network at steady state [3], subject to structural, reversibility, and flux capacity constraints [4], [5]. Structural constraints arise from the stoichiometry of the metabolic reactions. (Ir)reversibility constraints are thermodynamic in nature and capture the direction in which chemical substances flow within a reaction. Finally, flux capacity constraints can be derived from the availability of nutrients, the existence of a knockout, and biochemical data on the maximum throughput

This work is supported in part by the National Science Foundation under Grant CNS 1054604.

Michael M. Zavlanos is with the Dept. of Mechanical Engineering, Stevens Institute of Technology, Hoboken, NJ 07030, USA, michael.zavlanos@stevens.edu. A. Agung Julius is with the Dept. of Electrical, Computer and Systems Engineering, Rensselaer Polytechnic Institute, Troy, NY 12180, USA, agung@ecse.rpi.edu.

of enzymes. Given such constraints, the flux of chemical substrates through the network is limited to a feasible region defined by a convex polytope, and the objective of FBA is to determine a feasible set of fluxes that is best manifested in the biological system under consideration. The assumption commonly made is that the metabolic system exhibits a metabolic state that is optimal in terms of cellular growth [4], [6]. Cellular growth can be represented by accumulation of cellular biomass, which is composed of cellular metabolites. If the composition of the cellular biomass is known, cellular growth can be captured by an artificial biomass reaction involving metabolites at stoichiometries defined by their contribution in biomass composition. Therefore, calculation of the optimal growth rate and the corresponding metabolic fluxes can be posed as a Linear Program (LP) [1], [2].

To date, robustness analysis of metabolic networks has primarily focused on the response of the network to structural changes, such as gene knockouts or gene deletions, and has traditionally relied on “brute force” FBA applied to different knockout combinations [7], [8]. Minimal cutset algorithms for knockout experiment design were proposed in [9], [10] and were recently extended to arbitrarily large networks [11]. The related literature also includes mixed integer linear programming approaches [12], [13], as well as a convex relaxations that scale better with the network size [14].

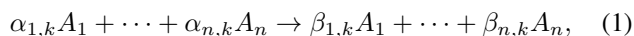
In this paper, we switch gears and study robustness of FBA to perturbations in the biomass composition and the metabolic flux capacities. These quantities are typically subject to measurement uncertainty, which raises the need for new Robust FBA (RFBA) techniques that return flux distributions that are optimal for worst case measurements. An important technical challenge in introducing uncertainty in FBA is that this typically appears in the stoichiometric equality constraints of the original LP, in the form of uncertain stoichiometric coefficients of an artificial biomass reaction [15], [16]. For this, we propose a least squares approximation of the original uncertain LP that results in a bi-criterion optimization problem. Using the S-procedure and tools from matrix analysis, we show that this approximation is equivalent to a semidefinite optimization problem that can be solved optimally using available techniques [17]. We illustrate our approach on synthetic data and study the effect of regularization on the final solution.

This paper is organized as follows: In Section II we describe FBA for maximization of cellular growth. In Section III we introduce measurement uncertainty and develop RFBA based on a least squares approximation of the original uncertain LP. We illustrate our approach on synthetic data in Section IV, and study the effect of regularization.

## II. FLUX BALANCE ANALYSIS (FBA)

### A. Metabolic Network Modeling at Steady State

Consider a metabolic network with  $n$  metabolites and  $m$  reactions. The  $k$ -th reaction can be written as



where  $A_i$  denotes the  $i$ -th metabolite, and  $\alpha_{\bullet,k}, \beta_{\bullet,k}$  are non-negative integers that denote the stoichiometric coefficients of the  $k$ -th reaction. Obviously, if  $A_i$  is not involved as a reactant in the  $k$ -th reaction, then  $\alpha_{i,k} = 0$ . Similarly, if  $A_i$  is not involved as a product in the  $k$ -th reaction, then  $\beta_{i,k} = 0$ . In regular reactions we have

$$\alpha_{\bullet,k} \neq 0, \quad (2a)$$

$$\beta_{\bullet,k} \neq 0, \quad (2b)$$

which means that there is always some reactant and product associated with the reaction. Here we assume that all reactions are irreversible. This is done without any loss of generality, since reversible reactions can be written as two opposite irreversible reactions.

In addition to the regular reactions, we also have uptake reactions. These are reactions that can be written as



and model the uptake of metabolite  $A_i$  from the environment. Uptake reactions can be also expressed as in (1), without the restriction of (2a).

If we denote the concentration of the  $i$ -th metabolite as  $x_i$  and the rate of the  $k$ -th reaction as  $\omega_k$ , then we can show that  $x$  and  $\omega$  are related through

$$\frac{dx}{dt} = (\beta - \alpha)\omega, \quad \omega \geq 0 \quad (4)$$

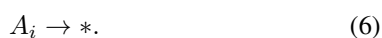
where  $\alpha$  and  $\beta$  are the  $n \times m$  matrices formed by the coefficients of (1), and the symbol  $\geq$  denotes element-wise inequality.

In microbes, the transient dynamics of the metabolic network are faster than both cellular growth rates and the dynamic changes in the organism's environment. In analyzing the network, thus, it is assumed that it is in its steady-state. In steady-state, the rates  $dx/dt$  represent the accumulation of metabolites and must be element-wise nonnegative. This is because the cell can act as a perpetual sink, but not as a perpetual source (without any uptake). Thus, in steady-state condition, the following relations hold:

$$(\beta - \alpha)\omega - \frac{dx}{dt} = 0, \quad (5a)$$

$$\omega \geq 0, \quad \frac{dx}{dt} \geq 0. \quad (5b)$$

We can rewrite (5) in a more compact form by introducing pseudo-reactions as sinks. These are reactions that can be written as



We associate a sink with every metabolite. Thus, there are  $n$  pseudo-reactions. Equation (5) can, therefore, be written compactly as [14]

$$Sv = 0, \quad v \geq 0, \quad (7)$$

where

$$S \triangleq [ \beta - \alpha \mid -I ] \in \mathbb{Z}_+^{n \times (m+n)}, \quad v \triangleq \begin{bmatrix} \omega \\ \frac{dx}{dt} \end{bmatrix} \in \mathbb{R}^{m+n}. \quad (8)$$

Since, typically, the number of reactions is greater than the number of metabolites, i.e.,  $S$  is a wide matrix, the system (7) may have multiple solutions corresponding to flux distributions representing different metabolic states. Therefore, the null space, or the set of all feasible flux distributions, represents the capabilities of the metabolic genotype. The transport fluxes represent environmental conditions that, along with the genotype, define the metabolic state. However, obtaining all possible metabolic states for any genotype-environment interaction depends on how well the genotype and environmental factors are characterized [18].

### B. Maximization of Cellular Growth

The objective of Flux Balance Analysis (FBA) is to determine a feasible metabolic state that is best manifested in the biological system under consideration. The assumption commonly made is that the metabolic system exhibits a metabolic state that is optimal under some criteria. In the case of cell growth, the objective is biomass production, i.e., the rate at which metabolic compounds are converted into biomass constituents, such as nucleic acids, proteins and lipids. Biomass production can be mathematically represented by an artificial biomass reaction [15], [16]



that consumes precursor metabolites  $A_i$  at stoichiometries  $b_i$  that simulate biomass production. The biomass reaction is based on experimental measurements of the biomass components  $b_i$  contained in the vector  $b$  and is scaled so that the flux through it is equal to the exponential growth rate  $\mu = \ln(2)/T$  of the organism, where  $T > 0$  is the doubling time. Reaction (9) introduces an additional column in the stoichiometric matrix, which becomes

$$S_b \triangleq [ \beta - \alpha \mid -b \mid -I ] \in \mathbb{R}_+^{n \times (n+m+1)}, \quad (10)$$

with corresponding flux vector

$$v_b \triangleq \left[ \omega^T \mid \omega_b \mid \frac{dx^T}{dt} \right]^T \in \mathbb{R}^{n+m+1}, \quad (11)$$

where  $\omega_b$  is the rate of the artificial biomass reaction (9). Therefore, we can define an optimization problem to determine the metabolic fluxes  $v_b$  that ensure desired cell growth, dictated by precursor requirements contained in  $b$ , as

$$\begin{aligned} & \text{maximize} && e_b^T v_b \\ & \text{subject to} && S_b v_b = 0 \\ & && 0 \leq v_b \leq v_{max} \end{aligned}, \quad (12)$$

where  $e_b$  is a column vector with all entries equal to zero except for the  $(m+1)$ -st entry that is equal to one and corresponds to the position of  $\omega_b$  in  $v_b$  (c.f. (11)). In problem (12), we have also included flux capacities  $v_{max}$ , which in the case of the precursors correspond to their actual experimentally measured concentrations for given cell growth. If no such knowledge is available, the fluxes can be unconstrained.

### III. ROBUST FLUX BALANCE ANALYSIS (RFBA)

The experimentally measured biomass composition vector  $b \in \mathbb{R}_+^n$  and the flux capacities  $v_{max} \in \mathbb{R}_+^{n+m+1}$  are typically subject to uncertainty. In this section we introduce measurement uncertainty in (12) and propose a reformulation of the FBA problem that is robust with respect to worst case parameter uncertainty.

Observe first that parameter uncertainty enters (12) in the equality constraints  $S_b v_b = 0$ , which poses technical difficulties in finding a unique flux distribution  $v_b$  that satisfies these constraints for all possible evaluations of  $b$  within an uncertainty set. Therefore, we approximate (12) by the least squares bi-criterion optimization problem

$$\begin{aligned} & \text{minimize} && \epsilon \|S_b v_b\|_2 - (1-\epsilon) e_b^T v_b, \\ & \text{subject to} && 0 \leq v_b \leq v_{max} \end{aligned} \quad (13)$$

where  $\epsilon \in [0, 1]$  is a tuning (regularization) parameter [17] that regulates the relative contribution of the two objectives  $\|S_b v_b\|_2$  and  $-e_b^T v_b$  in (13). In problem (13) we trade exact satisfaction of the stoichiometric equality constraints for maximization of biomass. In choosing  $\epsilon$ , we should ensure that equality violation is not too large, i.e., that the stoichiometric error  $\|S_b v_b\|_2$  is small enough. We will study sensitivity of the solution of problem (13) to the tuning parameter  $\epsilon$  in Section IV.

#### A. Uncertainty in the Biomass Composition

To model the uncertainty in the biomass composition vector  $b \in \mathbb{R}_+^n$ , assume that there are  $p$  available measurements  $\{b_i\}_{i=1}^p \in \mathbb{R}_+^n$  of the biomass composition and for every  $\xi \in \mathbb{R}^p$  with  $\|\xi\|_2 \leq \rho$  let

$$b(\xi) = b_0 + \sum_{i=1}^p \xi_i b_i$$

where  $b_0 = \frac{1}{p} \sum_{i=1}^p b_i$  denotes a mean biomass composition vector ( $b_0$  can also be taken the zero vector  $0^n$ ).<sup>1</sup> Then, the stoichiometric matrix  $S_b$  becomes

$$S_b(\xi) = S_{b_0} + \sum_{i=1}^p \xi_i S_{b_i},$$

<sup>1</sup>Hereafter,  $b_i$  will denote the  $i$ -th measurement of the biomass composition vector, rather than the stoichiometric coefficient of the  $i$ -th metabolite in the biomass reaction previously defined in (9). The stoichiometric coefficient of the  $j$ -th metabolite of the  $i$ -th measurement of the biomass composition vector will be denoted by  $b_{ij}$ .

with  $S_{b_i}$  defined as in (10) for biomass composition vector  $b_i$ . Therefore, we can define the robust counterpart of problem (13) by

$$\begin{aligned} & \text{minimize} && \epsilon r(S_b, v_b, \rho) - (1-\epsilon) e_b^T v_b \\ & \text{subject to} && 0 \leq v_b \leq v_{max} \end{aligned} \quad (14)$$

where

$$r(S_b, v_b, \rho) = \max_{\|\xi\|_2 \leq \rho} \|S_b(\xi) v_b\|_2 \quad (15)$$

denotes the worst case stoichiometric error. Without the presence of the objective  $e_b^T v_b$ , problem (14) is also known as a Robust Least Squares problem [19], [20]. Let

$$M(v_b) = [S_{b_1} v_b \quad \dots \quad S_{b_p} v_b]$$

and define the quantities

$$F = M^T(v_b) M(v_b), \quad g = M^T(v_b) S_{b_0} v_b, \quad h = \|S_{b_0} v_b\|_2^2.$$

Then,

$$\begin{aligned} \|S_b(\xi) v_b\|_2^2 &= \left\| S_{b_0} v_b + \sum_{i=1}^p \xi_i S_{b_i} v_b \right\|_2^2 \\ &= \|S_{b_0} v_b + [S_{b_1} v_b \quad \dots \quad S_{b_p} v_b] \xi\|_2^2 \\ &= (S_{b_0} v_b + M(v_b) \xi)^T (S_{b_0} v_b + M(v_b) \xi) \\ &= h + g^T \xi + \xi^T g + \xi^T F \xi \\ &= \begin{bmatrix} 1 & \xi^T \end{bmatrix} \begin{bmatrix} h & g^T \\ g & F \end{bmatrix} \begin{bmatrix} 1 \\ \xi \end{bmatrix}, \end{aligned}$$

which gives

$$r^2(S_b, v_b, \rho) = \max_{\|\xi\|_2 \leq \rho} \begin{bmatrix} 1 & \xi^T \end{bmatrix} \begin{bmatrix} h & g^T \\ g & F \end{bmatrix} \begin{bmatrix} 1 \\ \xi \end{bmatrix}.$$

Therefore, minimizing  $r(S_b, v_b, \rho)$  is equivalent to minimizing  $\lambda \geq 0$  such that

$$\begin{bmatrix} 1 & \xi^T \end{bmatrix} \begin{bmatrix} h & g^T \\ g & F \end{bmatrix} \begin{bmatrix} 1 \\ \xi \end{bmatrix} \leq \lambda$$

for all possible  $\xi \in \mathbb{R}^p$  with  $\xi^T \xi \leq \rho^2$ . In other words, we need to find a minimum scalar  $\lambda$  and a vector  $v_b$  such that

$$\begin{bmatrix} 1 & \xi^T \end{bmatrix} \begin{bmatrix} \lambda - h & -g^T \\ -g & -F \end{bmatrix} \begin{bmatrix} 1 \\ \xi \end{bmatrix} \geq 0$$

whenever

$$\begin{bmatrix} 1 & \xi^T \end{bmatrix} \begin{bmatrix} \rho^2 & 0 \\ 0 & -I \end{bmatrix} \begin{bmatrix} 1 \\ \xi \end{bmatrix} \geq 0,$$

for all  $\xi \in \mathbb{R}^p$ . By the S-procedure, this happens if and only if

$$\begin{bmatrix} \lambda - h & -g^T \\ -g & -F \end{bmatrix} \succeq \tau \begin{bmatrix} \rho^2 & 0 \\ 0 & -I \end{bmatrix},$$

for some  $\tau \geq 0$ .<sup>2</sup> Therefore, problem (14) can be equivalently written as

$$\begin{aligned} & \text{minimize} && \epsilon \lambda - (1-\epsilon) e_b^T v_b \\ & \text{subject to} && \begin{bmatrix} \lambda - \rho^2 \tau - h & -g^T \\ -g & \tau I - F \end{bmatrix} \succeq 0 \\ & && 0 \leq v_b \leq v_{max} \end{aligned} \quad (16)$$

<sup>2</sup>We write  $X \succeq 0$  if and only if the symmetric matrix  $X \in \mathbb{S}^n$  belongs in the positive semidefinite cone, defined by  $\mathbb{S}_+^n = \{X \in \mathbb{S}^n \mid X \succeq 0\}$ .

Since

$$\begin{aligned} & \begin{bmatrix} \lambda - \rho^2 \tau - \|S_{b_0} v_b\|_2^2 & -(M^T(v_b) S_{b_0} v_b)^T \\ -M^T(v_b) S_{b_0} v_b & \tau I - M^T(v_b) M(v_b) \end{bmatrix} \\ &= \begin{bmatrix} \lambda - \rho^2 \tau & 0 \\ 0 & \tau I \end{bmatrix} - \begin{bmatrix} (S_{b_0} v_b)^T \\ M^T(v_b) \end{bmatrix} I \begin{bmatrix} S_{b_0} v_b & M(v_b) \end{bmatrix}, \end{aligned}$$

we can apply Schur complements to problem (16) to obtain

$$\begin{aligned} & \text{minimize} && \epsilon \lambda - (1 - \epsilon) e_b^T v_b \\ & \text{subject to} && \mathcal{F}(\lambda, \tau, v_b) \succeq 0 \\ & && 0 \leq \rho^2 \tau \leq \lambda \\ & && 0 \leq v_b \leq v_{max} \end{aligned}, \quad (17)$$

where

$$\mathcal{F}(\lambda, \tau, v_b) = \begin{bmatrix} \lambda - \rho^2 \tau & 0 & (S_{b_0} v_b)^T \\ 0 & \tau I & M^T(v_b) \\ \hline S_{b_0} v_b & M(v_b) & I \end{bmatrix}. \quad (18)$$

Problem (17) is a semidefinite program in variables  $(\lambda, \tau, v_b)$  and can be solved using available techniques [17], [21].

### B. Uncertainty in the Metabolic Flux Capacities

To introduce uncertainty in the flux capacities  $v_{max} \in \mathbb{R}_+^{n+m+1}$ , assume that there are  $q$  available measurements  $\{v_{max}^k\}_{k=1}^q \in \mathbb{R}_+^{n+m+1}$ , and for every  $\zeta \in \mathbb{R}^q$  with  $\|\zeta\|_2 \leq \eta$  and every flux  $i = 1, \dots, n + m + 2$  let

$$e_i^T v_{max}(\zeta) = e_i^T v_{max}^0 + \sum_{k=1}^q \zeta_k e_i^T v_{max}^k,$$

where  $v_{max}^0 = \frac{1}{q} \sum_{k=1}^q v_{max}^k$  and  $e_i \in \mathbb{R}^{n+m+1}$  is a column vector with all entries equal to zero except for the  $i$ -th entry that is equal to one. The inner product of  $v_{max}$  with  $e_i$  corresponds to the capacity of the  $i$ -th flux. Therefore, (17) can be reformulated to account for uncertainty in the flux capacities as

$$\begin{aligned} & \text{minimize} && \epsilon \lambda - (1 - \epsilon) e_b^T v_b \\ & \text{subject to} && \mathcal{F}(\lambda, \tau, v_b) \succeq 0 \\ & && 0 \leq \rho^2 \tau \leq \lambda \\ & && 0 \leq e_i^T v_b \leq \inf_{\|\zeta\|_2 \leq \eta} \{e_i^T v_{max}(\zeta)\} \end{aligned}. \quad (19)$$

for all fluxes  $i = 1, \dots, n + m + 2$ . Let  $N = [v_{max}^1 \dots v_{max}^q]$  and observe that

$$\begin{aligned} \inf_{\|\zeta\|_2 \leq \eta} \{e_i^T v_{max}(\zeta)\} &= e_i^T v_{max}^0 + \inf_{\|\zeta\|_2 \leq \eta} \{e_i^T N \zeta\} \\ &= e_i^T v_{max}^0 + \eta \inf_{\|\zeta\|_2 \leq 1} \{e_i^T N(\zeta/\eta)\} \\ &= e_i^T v_{max}^0 + \eta \inf_{\|\zeta\|_2 \leq 1} \{e_i^T N \zeta\} \\ &= e_i^T v_{max}^0 - \eta \sup_{\|\zeta\|_2 \leq 1} \{-e_i^T N \zeta\} \\ &= e_i^T v_{max}^0 - \eta \|N^T e_i\|_*, \end{aligned}$$

where  $\|u\|_* = \sup\{u^T x \mid \|x\|_2 \leq 1\}$  denotes the dual norm of  $u$ , which can be interpreted as the operator norm of  $z^T$  if

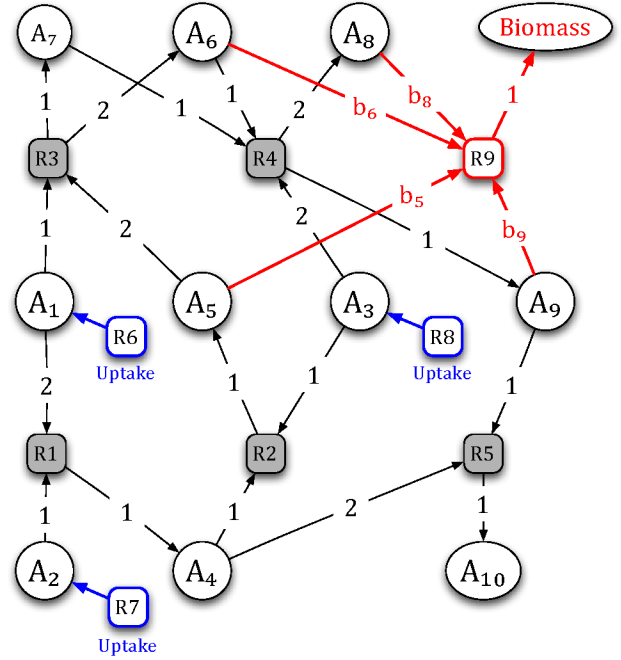


Fig. 1. Metabolic network consisting of 10 metabolites, 5 reactions, and one artificial biomass reaction. Shown are also reaction stoichiometries. Arrows indicate reaction direction one of the two reaction directions, considered positive. The artificial biomass reaction is indicated in red color, while the uptake reactions are shown in blue.

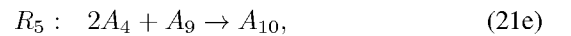
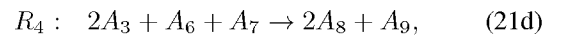
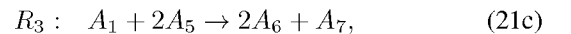
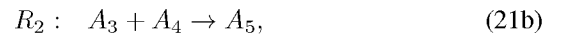
it is considered a  $1 \times n$  matrix.<sup>3</sup> Substituting in (19) we get

$$\begin{aligned} & \text{minimize} && \epsilon \lambda - (1 - \epsilon) e_b^T v_b \\ & \text{subject to} && \mathcal{F}(\lambda, \tau, v_b) \succeq 0 \\ & && 0 \leq \rho^2 \tau \leq \lambda \\ & && 0 \leq v_b \leq v_{max}^0 - \eta \text{vec}(\{\|N^T e_i\|_*\}_{i=1}^{n+m+2}) \end{aligned}, \quad (20)$$

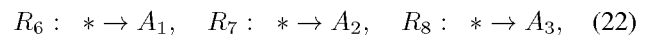
where  $\text{vec}(\{x_1, \dots, x_k\})$  denotes a column vector with elements  $x_1, \dots, x_k$ . Problem (20) is a semidefinite program and can be solved using available techniques [17], [21].

### IV. SIMULATION RESULTS

In this section, we illustrate problem (20) on synthetic metabolic networks and study the effect of regularization (tuning parameter  $\epsilon$ ) on the final solution. In particular, consider a metabolic network consisting of  $n = 10$  metabolites labeled  $\{A_i\}_{i=1}^{10}$ , and  $m = 5$  reversible reactions



three irreversible uptake reactions



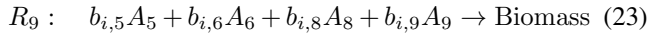
<sup>3</sup>Note that  $\inf\{X\} = -\sup\{-X\}$  for any set  $X$ , where  $-X = \{-x \mid x \in X\}$ .

TABLE I

STOICHIOMETRIC MATRIX  $[\beta - \alpha \mid -b]$  FOR THE METABOLIC NETWORK SHOWN IN FIG. 1. THE REACTIONS  $R_{\bullet}^{-}$  INDICATE OPPOSITE DIRECTIONALITY WITH RESPECT TO THE REACTIONS  $R_{\bullet}$ , AND ARE INTRODUCED TO MODEL REVERSIBILITY (21). REACTIONS  $R_6$  THROUGH  $R_8$  MODEL UPTAKE OF METABOLITES FROM THE ENVIRONMENT (22), WHILE REACTION  $R_9$  CORRESPONDS TO THE ARTIFICIAL BIOMASS REACTION (23).

	$R_1$	$R_2$	$R_3$	$R_4$	$R_5$	$R_1^{-}$	$R_2^{-}$	$R_3^{-}$	$R_4^{-}$	$R_5^{-}$	$R_6$	$R_7$	$R_8$	$R_9$
$A_1$	-2	0	-1	0	0	2	0	1	0	0	1	0	0	0
$A_2$	-1	0	0	0	0	1	0	0	0	0	0	1	0	0
$A_3$	0	-1	0	-2	0	0	-1	0	-2	0	0	0	1	0
$A_4$	1	-1	0	0	-2	-1	1	0	0	2	0	0	0	0
$A_5$	0	1	-2	0	0	0	-1	2	0	0	0	0	0	$-\mu b_5$
$A_6$	0	0	2	-1	0	0	0	-2	1	0	0	0	0	$-\mu b_6$
$A_7$	0	0	1	-1	0	0	0	-1	1	0	0	0	0	0
$A_8$	0	0	0	2	0	0	0	0	-2	0	0	0	0	$-\mu b_8$
$A_9$	0	0	0	1	-1	0	0	0	-1	1	0	0	0	$-\mu b_9$
$A_{10}$	0	0	0	0	1	0	0	0	0	-1	0	0	0	0

and an artificial biomass reaction



with  $b_i = [0 \ 0 \ 0 \ 0 \ b_{i,5} \ b_{i,6} \ 0 \ b_{i,8} \ b_{i,9} \ 0]^T$  the  $i$ -th measurement of the biomass composition vector, for  $i = 1, \dots, p$  with  $p = 10$  (Fig. 1). We assume that  $b_i$  has mean  $[0 \ 0 \ 0 \ 0 \ 3 \ 1 \ 0 \ 2 \ 1 \ 0]^T$  and that every one of its entries is subject to zero mean and 0.5 variance gaussian noise. Every biomass composition vector  $b_i$  is normalized so that  $b_{i,5} + b_{i,6} + b_{i,8} + b_{i,9} = 1$ . Let  $T = 1\text{h}$  be the doubling time of the organism, so that the growth rate is  $\mu = \ln(2)/T = 0.69\text{h}^{-1}$ . Then, the stoichiometric matrix of the network under consideration is shown in Table I. Furthermore, we assume that all fluxes are unconstrained, except for the uptake fluxes of reactions  $R_6$ ,  $R_7$  and  $R_8$  that are upper bounded by 0.1. For simplicity, we assume that these bounds are deterministic.

We evaluated the performance problem (20) for different values of the tuning parameter  $\epsilon \in [0, 1]$  and for  $\rho = 1.5$ . Simulations were performed in MATLAB using the `cvx` toolbox for disciplined convex programming [21]. Fig. 2 illustrates as a function of the tuning parameter  $\epsilon \in [0, 1]$  (a) the biomass objective  $e_b^T v_b$ , (b) the upper bound  $\lambda$  on the worst case stoichiometric error  $r(S_b, v_b, \rho)$  defined in (15), and (c) the stoichiometric errors  $\|S_{b_i} v_b\|_2$  for all measured biomass compositions  $b_i$ , with  $i = 1, \dots, p$ . Observe that the values of  $\|S_{b_i} v_b\|_2$  are always upper bounded by  $\lambda$ , since  $\rho = 1.5 \geq 1$ . In other words, the set  $\{\|S_b(\xi)v_b\|_2 \mid \|\xi\|_2 \leq \rho\}$  includes the errors  $\|S_{b_i} v_b\|_2$  for all measured biomass compositions (15). This is not necessarily the case if  $\rho < 1$ .

For  $\epsilon = 0$  the stoichiometric equality constraints are ineffective and, therefore, the resulting value of  $e_b^T v_b$  has no biological meaning. We are interested in regions of the plot where  $\lambda$  is minimum and  $e_b^T v_b$  is maximum. Note that  $\lambda$  can not become identically zero, since there does not exist a unique nontrivial flux vector  $v_b$  for which  $\|S_b v_b\|_2 = 0$  for all biomass compositions  $b \in \{b_0 + \sum_{i=1}^p \xi_i b_i \mid \|\xi\|_2 \leq \rho\}$ . From Fig. 2 we see that  $\lambda$  is almost at its minimum for  $\epsilon \geq 0.85$ . Since  $e_b^T v_b$  decreases rapidly as  $\epsilon$  increases beyond 0.85, we choose  $\epsilon = 0.85$  to obtain  $e_b^T v_b = 0.1593$ . The flux vector  $v_b$  obtained by the solution of problem (20) for

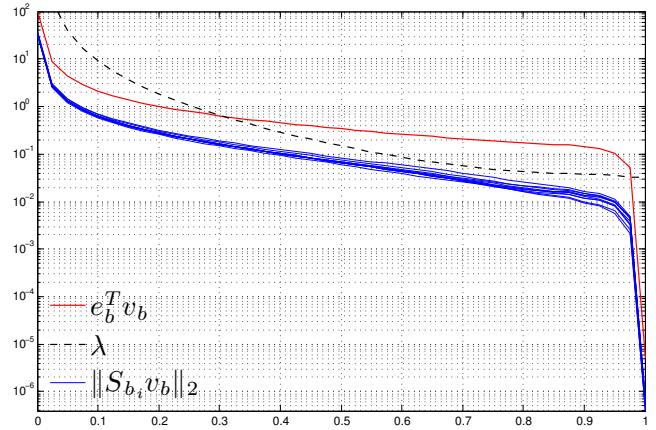


Fig. 2. Plots of (a) the biomass objective  $e_b^T v_b$ , (b) the upper bound  $\lambda$  on the worst case stoichiometric error  $r(S_b, v_b, \rho)$  defined in (15), and (c) the stoichiometric errors  $\|S_{b_i} v_b\|_2$  for all measured biomass composition vectors  $b_i$ , with  $i = 1, \dots, p$ , with respect to the tuning parameter  $\epsilon \in [0, 1]$ . The  $y$ -axis is in log-scale.

$\epsilon = 0.85$  is shown in Table II. Note that for this value of  $\epsilon = 0.85$  there is maximum uptake of metabolites  $A_1$  and  $A_3$  (fluxes  $\omega_6$  and  $\omega_8$ ) equal to the upper bound 0.1, which is distributed among the five reactions to result in metabolites that maximize biomass. All steady state metabolite concentrations are effectively constant ( $dx_i/dt = 0$ ), except for the concentrations of  $A_2$  and  $A_9$  that slightly increase. This increase is due to the stoichiometries in the network that prevent these metabolites from being fully consumed to produce biomass. The values of the errors  $\|S_{b_i} v_b\|_2$  for  $i = 1, \dots, p$  are small, ranging between 0.01–0.02 in value.

*Remark 4.1* ( $\epsilon \rightarrow 1$ ): Observe that as  $\epsilon \rightarrow 1$ , both  $e_b^T v_b \rightarrow 0$  and  $\|S_{b_i} v_b\|_2 \rightarrow 0$ , for all measurements  $i = 1, \dots, p$  (Fig. 2). This behavior is justified, since as  $\epsilon \rightarrow 1$  the biomass objective  $e_b^T v_b$  becomes effectively inactive and the optimization problem (20) determines fluxes  $v_b$  to only minimize the worst case stoichiometric error  $r(S_b, v_b, \rho)$ . Numerically, a *sparse* flux vector  $v_b$  as the one shown in Table II is not optimal as a minimizer of  $r(S_b, v_b, \rho)$ . In fact, if  $\epsilon = 1$ , the flux vector  $v_b$  returned by (20) for the metabolic network of Fig. 1 is rather *dense* with

TABLE II

FLUX VECTOR  $v_b$  DETERMINED BY PROBLEM (20) FOR THE NETWORK ILLUSTRATED IN FIG. 1 AND FOR TUNING PARAMETER  $\epsilon = 0.85$ . THE FLUXES  $\omega_1$  THROUGH  $\omega_b$  ARE POSITIVE IF THEIR DIRECTION AGREES WITH REACTIONS (21), (22) AND (23), AS SHOWN IN FIG. 1.

Flux	Value	Flux	Value
$\omega_1 = \omega(R_1)$	0.0448	$dx_1/dt$	0.0000
$\omega_2 = \omega(R_2)$	0.0701	$dx_2/dt$	0.0315
$\omega_3 = \omega(R_3)$	0.0129	$dx_3/dt$	0.0000
$\omega_4 = \omega(R_4)$	0.0156	$dx_4/dt$	0.0000
$\omega_5 = \omega(R_5)$	-0.0101	$dx_5/dt$	0.0000
$\omega_6 = \omega(R_6)$	0.1000	$dx_6/dt$	0.0000
$\omega_7 = \omega(R_7)$	0.0764	$dx_7/dt$	0.0000
$\omega_8 = \omega(R_8)$	0.1000	$dx_8/dt$	0.0000
Biomass Reaction		$dx_9/dt$	0.0081
$\omega_b = \omega(R_9)$	0.1593	$dx_{10}/dt$	0.0000

TABLE III

FLUX VECTOR  $v_b$  DETERMINED BY PROBLEM (20) FOR THE NETWORK ILLUSTRATED IN FIG. 1 AND FOR TUNING PARAMETER  $\epsilon = 1.00$ . THE FLUXES  $\omega_1$  THROUGH  $\omega_b$  ARE POSITIVE IF THEIR DIRECTION AGREES WITH REACTIONS (21), (22) AND (23), AS SHOWN IN FIG. 1.

Flux	Value	Flux	Value
$\omega_1 = \omega(R_1)$	0.0327	$dx_1/dt$	0.0131
$\omega_2 = \omega(R_2)$	0.0234	$dx_2/dt$	0.0400
$\omega_3 = \omega(R_3)$	0.0091	$dx_3/dt$	0.0383
$\omega_4 = \omega(R_4)$	0.0057	$dx_4/dt$	0.0058
$\omega_5 = \omega(R_5)$	0.0018	$dx_5/dt$	0.0052
$\omega_6 = \omega(R_6)$	0.0876	$dx_6/dt$	0.0125
$\omega_7 = \omega(R_7)$	0.0727	$dx_7/dt$	0.0034
$\omega_8 = \omega(R_8)$	0.0730	$dx_8/dt$	0.0114
Biomass Reaction		$dx_9/dt$	0.0039
$\omega_b = \omega(R_9)$	$5.27 \cdot 10^{-6}$	$dx_{10}/dt$	0.0018

very little biomass is produced (Table III). As expected, all resources are consumed to produce metabolites  $A_1, \dots, A_{10}$  and obtain a dense flux vector  $v_b$ . The stoichiometric errors  $\{\|S_b v_b\|_2\}_{i=1}^p$  in this case are of the order of  $10^{-7}$ , as expected (but still nonzero).

## V. CONCLUSIONS

Metabolic Flux Balance Analysis (FBA) is a powerful optimization-based technique that studies the feasible and optimal reaction fluxes through the network at steady state, subject to structural, reversibility, and flux capacity constraints. Among the large number of possible flux distributions, FBA determines the one that is best manifested in the system under consideration. The assumption commonly made is that the metabolic systems exhibits a metabolic state that is optimal in terms of cellular growth, which is typically represented by accumulation of cellular biomass.

In this paper, we proposed Robust Flux Balance Analysis (RFBA) to account for uncertainty in the biomass composition and flux capacities. We showed that flux distributions that are robust to worst case parameter uncertainty can

be obtained by the solution of a bi-criterion semidefinite program, which can be solved to optimality using available techniques and scales well to large networks due its convex nature. We illustrated our approach on synthetic data and studied the effect of regularization on the final solution. Future work involves application of our method to real experimental data and comparison with existing techniques.

## REFERENCES

- [1] A. Varma and B. O. Palsson, "Metabolic flux balancing: Basic concepts, scientific and practical use," *Nature Biotechnology*, vol. 12, pp. 994–998, 1994.
- [2] J. S. Edwards, M. W. Covert, and B. O. Palsson, "Metabolic modeling of microbes: The flux balance approach," *Environmental Microbiology*, vol. 4, no. 3, pp. 133–140, 2002.
- [3] U. Alon, *An Introduction to Systems Biology*. Chapman and Hall, 2007.
- [4] N. D. Price, J. L. Reed, and B. O. Palsson, "Genome-scale models of microbial cells: evaluating the consequences of constraints," *Nature Reviews Microbiology*, vol. 2, no. 1, pp. 886–897, 2004.
- [5] J. L. Reed, T. D. Vo, C. H. Schilling, and B. O. Palsson, "An expanded genome-scale model of escherichia coli k-12," *Genome Biology*, vol. 4, no. 9, 2003.
- [6] A. Varma and B. O. Palsson, "Stoichiometric flux balance models quantitatively predict growth and metabolic by-product secretion in wild-type escherichia coli w3110," *Applied Environmental Microbiology*, vol. 60, pp. 3724–3731, 1994.
- [7] I. Thiele, T. D. Vo, N. D. Price, and B. O. Palsson, "Expanded metabolic reconstruction of Helicobacter pylori (iT341 GSM/GPR): an in silico genome-scale characterization of single- and double-deletion mutants," *Journal of Bacteriology*, vol. 187, pp. 5818–5830, 2005.
- [8] D. Deutscher, I. Meilijson, M. Kupiec, and E. Ruppin, "Multiple knockout analysis of genetic robustness in the yeast metabolic network," *Nature Genetics*, vol. 38, pp. 993–998, 2006.
- [9] S. Klant and E. D. Gilles, "Minimal cut sets in biochemical reaction networks," *Bioinformatics*, vol. 20, pp. 226–234, 2004.
- [10] S. Klant, "Generalized concept of minimal cut sets in biochemical networks," *Biosystems*, 2005.
- [11] M. Imielinski and C. Belta, "Exploiting the pathway structure of metabolism to reveal high-order epistasis," *BMC Systems Biology*, vol. 2, no. 40, 2008.
- [12] A. P. Burgard and C. D. Maranas, "Probing the performance limits of the escherichia coli metabolic network subject to gene additions or deletions," *Biotechnology and Bioengineering*, vol. 74, pp. 364–375, 2001.
- [13] A. P. Burgard, P. Pharkya, and C. D. Maranas, "Optknock: a bilevel programming framework for identifying gene knockout strategies for microbial strain optimization," *Biotechnology and Bioengineering*, vol. 84, pp. 647–657, 2003.
- [14] A. A. Julius, M. Imielinski, and G. J. Pappas, "Metabolic networks analysis using convex optimization," in *Proc. 47th IEEE Conference on Decision and Control*, Cancun, Mexico, December 2008, pp. 762–767.
- [15] J. S. Edwards and B. O. Palsson, "Metabolic flux balance analysis and the in silico analysis of escherichia coli k-12 gene deletions," *BMC Bioinformatics*, vol. 1, no. 1, pp. 695–704, 2000.
- [16] J. D. Orth, I. Thiele, and B. O. Palsson, "What is flux balance analysis?" *Nature Biotechnology*, vol. 28, no. 3, pp. 245–248, 2010.
- [17] S. P. Boyd and L. Vandenberghe, *Convex Optimization*. Cambridge University Press, 2004.
- [18] R. Ramakrishna, J. S. Edwards, A. McCulloch, and B. O. Palsson, "Flux-balance analysis of mitochondrial energy metabolism: Consequences of systemic stoichiometric constraints," *American Journal of Physiology – Regulatory, Integrative and Comparative Physiology*, vol. 280, pp. 695–704, 2001.
- [19] A. Ben-Tal, L. El Ghaoui, and A. Nemirovski, *Robust Optimization*. Princeton University Press, 2009.
- [20] L. El Ghaoui and H. Lebret, "Robust solutions to least squares problems with uncertain data," *SIAM: SIAM Journal on Matrix Analysis and Applications (SIMAX)*, vol. 18, no. 4, pp. 1035–1064, 1997.
- [21] S. P. Boyd and M. C. Grant, "cvx – MATLAB software for disciplined convex programming," 2005, <http://www.stanford.edu/~boyd/cvx/>.